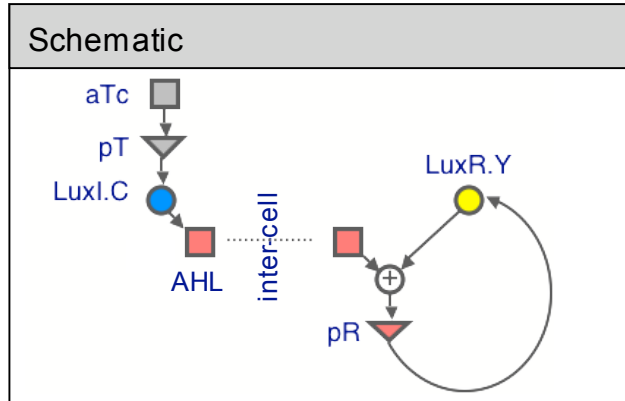


[Sen-TIC+Rec-RRY] Closed-Loop System

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	Host: Ecoli K12Z1
	p1.lacI::p2.tetR
	Parts: [Sen-TIC+Rec-RRY]
	pT.luxI.C
	pR.luxR.Y
	Small molecules
	Promoters
	Proteins
	Description
	Usage and compatibility

	Characteristics															
<p>Protocol: We grew sender cells overnight in LB, then transferred them to Glu-M9 minimal medium containing the desired final concentration of aTc, and allowed them to grow for ~12h, until they reached OD600=0.2. We then removed the sender cells by filtration, retaining the medium (which now contains AHL) and replenishing it with an equal volume of fresh Glu-M9 with double the standard glucose concentration. Receiver cells, which had previously been grown overnight in LB, were added to this medium. These cells were allowed to grow for ~12h, and harvested at OD600 < 0.1. Both sender and receiver cells were concentrated by centrifugation, and imaged on an epifluorescence microscope. We calculated the fluorescence per unit area of single cells, obtaining data from ~500 cells for each run. We then averaged these values in log space to obtain the final estimate of protein expression. The aTc mesh was [0 1 5 10 20 50] ng/ml (shown below the datapoints). <i>The results shown below are preliminary.</i></p>																
<table border="1"> <caption>Data points from the scatter plot</caption> <thead> <tr> <th>aTc concentration (ng/ml)</th> <th>log10(LuxI.C)</th> <th>log10(LuxR.Y)</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>~2.2</td> <td>~3.2</td> </tr> <tr> <td>5</td> <td>~2.4</td> <td>~3.2</td> </tr> <tr> <td>10</td> <td>~3.2</td> <td>~3.4</td> </tr> <tr> <td>50</td> <td>~3.4</td> <td>~4.1</td> </tr> </tbody> </table>		aTc concentration (ng/ml)	log10(LuxI.C)	log10(LuxR.Y)	0	~2.2	~3.2	5	~2.4	~3.2	10	~3.2	~3.4	50	~3.4	~4.1
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<p>Measurements and analysis were carried out by members of the NCBS iGEM 2007 team: Kiran, Krishna, Mukund, Navneet, Nilesch, Senthil, Shashanka, Sugat, Sushant, Varun, Vini, Vivek</p>																

•aTc: anhydrotetracycline.

•AHL: acyl homoserine lactone.

•p1: Constitutive LacIq promoter.

•p2: Constitutive N25 promoter.

•pT: Tet promoter

•pR: *V. fischeri* LuxpR promoter.

•LacI: Lac repressor, negative regulator of pL.

•TetR: Tet repressor, negative regulator of pT.

•LuxI: *V. fischeri* LuxI protein, synthesizes AHL.

•LuxR: *V. fischeri* LuxR protein, positive regulator of pR.

•C: Cyan fluorescent protein

•Y: Yellow fluorescent protein

Sender: aTc drives the expression of the LuxI protein (monitored using polycistronic CFP). This in turn drives the synthesis of AHL, which is exported. Receiver: In the presence of AHL, the LuxR protein activates its own expression at promoter pR (monitored using polycistronic YFP).

This is a stand-alone system.